



Fig. 2. Domain architecture of proteins containing the BSD domain. Only proteins with distinct modular organizations are shown. The domain names are according to those in the Simple Modular Architecture Research Tool [13] (<http://smart.embl-heidelberg.de>). Note that the name synapse-associated protein might be misleading as synaptic localization has so far only been demonstrated for *Drosophila melanogaster* [4], and close homologues are found in species without synapses. Abbreviations: BSD, found in BTF2-like transcription factors, synapse-associated and DOS2-like proteins; BTB, Broad-complex, Tamtrack and Bric a Brac; Ubox, a modified Ring finger domain associated with ubiquitination.

generated [8] for all candidates to identify the potential domain boundaries (Fig. 1). From this, the BSD domain appears to be ~60 amino acids in length. Secondary

structure prediction with PHD [9] indicates the presence of three α helices, which probably form a three-helical bundle in small domains. The third predicted helix

contains neighbouring phenylalanine and tryptophan residues – less common amino acids that are invariant in all the BSD domains identified and that are the most striking sequence features of the domain (Fig. 1). The BSD domain is found in a variety of species from primal protozoan to human, indicating a conserved, probably important, function.

Although the BSD domain occurs in very different protein families (e.g. synapse-associated proteins, hypothetical proteins and transcription factors), the presence of the novel domain in transcription factors suggests a role in chromatin-associated processes. The domain architectures of additional BSD domain-containing proteins are consistent with this assumption, but also make other functions feasible. For example, the BSD domain can co-occur with Ubox domains (Fig. 2), which are known to be involved in ubiquitination [6]. Although several proteins involved in the ubiquitination process are known to be associated with chromatin [10,11], this is not a prerequisite. BSD domains can also precede a BTB domain (Fig. 2), a protein–protein interaction domain that frequently occurs in transcription factors, in which it is succeeded by ZnF_C2H2 DNA-binding domains [7,12]. These findings suggest that the BSD domain could have a role in DNA binding, although it should be noted that neither synapse-associated proteins nor DOS2-like proteins are known to be associated with chromatin.

In summary, the delineation of the BSD domain and its boundaries (Fig. 1) should allow directed structural studies to test the involvement of this domain in chromatin-associated or more general processes.

References

- Fischer, L. *et al.* (1992) Cloning of the 62-kilodalton component of basic transcription factor BTF2. *Science* 257, 1392–1395
- Gileadi, O. *et al.* (1992) Cloning of a subunit of yeast RNA polymerase II transcription factor b and CTD kinase. *Science* 257, 1389–1392
- Altschul, S.F. *et al.* (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs *Nucleic Acids Res.* 25, 3389–3402
- Reisch, D. *et al.* (1995) The sap47 gene of *Drosophila melanogaster* codes for a novel conserved neuronal protein associated with synaptic terminals. *Brain Res. Mol. Brain Res.* 32, 45–54
- Eddy, S.R. (1998) Profile hidden Markov models. *Bioinformatics* 14, 755–763
- Aravind, L. and Koonin, E.V. (2000) The U box is a modified RING finger – a common domain in ubiquitination. *Curr. Biol.* 10, R132–R134

Box 1. Artificial *in silico* support of function prediction*

Currently, one of the proteins in the alignment, DOS1, is annotated in most databases to be involved in single-copy DNA replication and ubiquitination. This assumed function would match our findings that the BSD domain is present in basal transcription factors and could have a role in DNA-binding.

The functional description is based on a mutation in a region around the yeast open reading frame YDR068W (dating back to 1995), and the gene was originally named DOS1 in *Saccharomyces cerevisiae*. Further studies revealed that the mutation is localized in an adjacent gene; therefore, YDR068w was renamed DOS2 in GenBank at a later stage, without functional description. However, in the meantime, the old name and functional implications of the gene were imported into different public databases, where the erroneous entry can often still be found (Table I).

Table I. Variable nomenclature found in databases

Date of information retrieval	Correct database entries	Erroneous database entries	Erroneous database entry
	Gene names: DOS2, YDR068W, YD9609.22, YD8554.01, D4267 Function: hypothetical	Wrong gene names: DOA4, DOS1, UBP4, SSV7, NPI2, YDR069C	Correct gene name, but functional prediction of DOS1
September–October 2001	GenBank: AAA66522 NP_010353 EMBL: CAB16584 (one representative)	Sptrembl: O13905 Q9P5L4 SWISSPROT: P54858 EMBL: CAB91683 PIR: S54052 T49702	
28 November 2001	GenBank: AAA66522 NP_010353 EMBL: CAB16584 (one representative)	Sptrembl: O13905 Q9P5L4 EMBL: CAB91683 PIR: S54052 T49702	SWISSPROT: P54858

*Tim Formosa (University of Utah, Salt Lake City, UT 84112-5330, USA) was a coauthor of Box 1.

- 7 Zollman, S. *et al.* (1994) The BTB domain, found primarily in zinc finger proteins, defines an evolutionarily conserved family that includes several developmentally regulated genes in *Drosophila*. *Proc. Natl. Acad. Sci. U. S. A.* 91, 10717–10721
- 8 Thompson, J.D. *et al.* (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 22, 4673–4680
- 9 Rost, B. *et al.* (1994) PHD an automatic mail server for protein secondary structure prediction *CABIOS* 10, 53–60
- 10 Hershko, A. and Ciechanover, A. (1998) The ubiquitin system. *Annu. Rev. Biochem.* 67, 425–479
- 11 Ciechanover, A. (1998) The ubiquitin–proteasome pathway: on protein death and cell life. *EMBO J.* 17, 7151–7160
- 12 Bardwell, V.J. and Treisman, R. (1994) The POZ domain: a conserved protein–protein interaction motif. *Genes Dev.* 15, 1664–1677
- 13 Schultz, J. *et al.* (2000) SMART: a web-based tool for the study of genetically mobile domains. *Nucleic Acids Res.* 28, 231–234

Tobias Doerks*

Peer Bork

EMBL, 69012 Heidelberg, Meyerhofstr. 1, and Max-Delbrueck-Centrum, Berlin, Germany.

*e-mail: doerks@embl.heidelberg.de

Saskia Huber

Erich Buchner

Lehrstuhl für Genetik und Neurobiologie, Biozentrum der Universität, Am Hubland, 97074 Würzburg, Germany.

Sec61 β – a component of the archaeal protein secretory system

Lisa N. Kinch, Milton H. Saier, Jr and Nick V. Grishin

Sec61p/SecYEG complexes mediate protein translocation across membranes and are present in both eukaryotes and bacteria. Whereas homologues of Sec61 α /SecY and Sec61 γ /SecE exist in archaea, identification of the third component (Sec61 β or SecG) has remained elusive. Using PSI-BLAST, the archaeal counterpart of Sec61 β has been detected. With the identification of the Sec61 β motif, functions for a universal family of archaeal proteins can be predicted and the archaeal translocon system can be definitively detected.

Detection and rationalization of motifs in membrane proteins are more difficult than in soluble proteins because of their biased amino acid composition that is restricted to mostly hydrophobic residues and to a limited number of available spatial structures. The single transmembrane-spanning protein Sec61 β (Sbh1p in yeast) interacts with two other integral membrane proteins (Sec61 α and Sec61 γ) to form the core of the eukaryotic protein translocation machinery (reviewed in Refs [1,2]). The bacterial counterpart of this machine consists of a similar complex (SecYEG), with SecY and SecE representing homologues of Sec61 α and Sec61 γ , respectively [3]. The third bacterial membrane protein, SecG, differs somewhat from Sec61 β in both the number of membrane-spanning regions and residue conservation. This divergence brings into question the evolutionary origins of this third subunit, although SecG and Sec61 β both function to stimulate protein translocation activities and are thought to

be homologous [4,5]. Although archaeal homologues of SecY/Sec61 α and SecE/Sec61 γ exist, the identification of an archaeal homologue to either Sec61 β or SecG has remained elusive. The Sec61p/SecYEG system is universally present in all eukaryotes and bacteria for which completely sequenced genomes are available (T. Cao and M.H. Saier, Jr, unpublished). Thus, the absence of a Sec61 β /SecG homologue in archaea is puzzling. Based on sequence analyses, we have identified the third component of the archaeal translocation machinery. The archaeal counterpart resembles eukaryotic Sec61 β , suggesting an overall functional similarity between the translocation apparatus of archaea and the eukaryotes. Although this functional similarity awaits experimental confirmation, it mimics similarities displayed in other universal processes such as DNA replication, transcription and translation [6], and provides additional data for the studies of archaeal evolutionary origin [7].

We first detected a possible archaeal counterpart (gi | 15920503) to the human Sec61 β sequence (gi | 5803165) using PSI-BLAST [8] (parameters described in Fig. 1). Upon searching protein databases for related archaeal sequences, we found hits in all but two of the completely sequenced archaeal genomes. Searches against the nucleotide databases of these genomes suggested that these sequences (AE000914 and AE006662) were missed in gene prediction efforts. To substantiate the link to eukaryotic Sec61 β sequences, we generated a position-specific scoring matrix with a multiple sequence alignment

of the archaeal Sec61 β . Using this matrix, we initiated PSI-BLAST searches with each sequence from the alignment as a query. Two archaeal sequences used as queries (gi | 15920503 or RAP00437) identified the eukaryotic Sec61 β sequence (gi | 15239337) with significant statistics (E-value 0.002). This E-value, representing the estimated number of alignments with scores no less than that of a given alignment that is expected to occur in a database search by chance [8], falls below the threshold observed for distant homologues (E = 0.01) [9].

The short 45-residue motif identifying Sec61 β consists of a single, mostly hydrophobic stretch of ~20 amino acids preceded by a region of similar size that starts with several small amino acids and displays a particular residue conservation pattern (Fig. 1). The hydrophobic segment is predicted to form a transmembrane helix, with the C terminus of the helix defined by small and positively charged residues. The sequence of the helix incorporates a small residue at the beginning of the third turn and a relatively conserved histidine in the last turn. We suggest that the most conserved residue in the motif (proline) forms part of the N-terminal cap structure of this helix. The sequence between this helix and the stretch of small residues at the N terminus of the motif is characterized by four predominantly charged positions, having two negative charges surrounded by positive charges on either side. The archaeal sequences additionally contain conserved positively charged residues N-terminal to the transmembrane helix,