Protein Sequence Motif

BSD: a novel domain in transcription factors and synapse-associated proteins

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This article describes a novel domain, BSD, that is present in basal transcription factors, synapse-associated proteins and several hypothetical proteins. It occurs in a variety of species ranging from primal protozoan to human. The BSD domain is characterized by three predicted α helices, which probably form a three-helical bundle, as well as by conserved tryptophan and phenylalanine residues, located at the C terminus of the domain.

Initiation of transcription by **RNA-polymerase B requires several** accessory proteins. Mammalian BTF2 and its yeast homologue TFB1 are essential transcription factors in this initiation event [1,2]. The modular architecture of these proteins is not well understood: no functional details nor domains are characterized. We have analysed different regions of BTF2 to identify possible functional domains. PSI-BLAST [3] searches against the NRDB (non-redundant database) with a central region (conserved residues 180-232, Fig. 1) of BTF2 reveal weak similarity (E value = 0.14) to hypothetical DOS2-like proteins (Box 1) and to a family of proteins with homology (E value = 2.4) to a synapse-associated protein of Drosophila melanogaster.

Synapse-associated proteins are expressed specifically in neurons and act as important molecular elements of the nervous system [4]. The similarities of BTF2 to these proteins are just below the default thresholds used by BLAST and are thus indicative of homology, although the significance remains to be proven. Reciprocal PSI-BLAST and PHI-BLAST searches reveal lower E values (e.g. E value = 0.11 using DOS2 as query), although these are still not significant. However, statistically significant similarity is shown by using reciprocal Hidden Markov Model (HMMer) searches [5] as an independent method, starting with the synapse-associated protein family and its related hypothetical plant homologues (including a Ubox-domain-containing

TFB1_a	sc	165	LDDSLSKEKLLTNLKLQQSLLKGNKVLMKVFQETVINAGLPPSEFWSTRIPLLRAFA	P32776
TFB1_b	sc	243	SENKVNVNLSREKILNIFENYPIVKKAYTDNVPKNFKEPEFWARFFSSKLFRK	P32776
BTF2_a	hs	99	LLPKFKRKANKELEEKNRMLQEDPVLFQLYKDLVVSQVISAEEFWANRLNVNATDS	P32780
BTF2_b	hs	180	GCNGLRYNLTSDIIESIFRTYPAVKMKYAENVPHNMTEKEFWTRFFQSHYFHR	P32780
TFB1dm_a	dm	109	LLPNFKRKVDKDLEDKNRILVENPNLLQLYKDLVITKVLTSDEFWATHAKDHALKK	Q9V713
TFB1dm_b	dm	182	GCNGLKYNLTSDVIHCIFKTYPAVKRKHFENVPAKMSEAEFWTKFFQSHYFHR	Q9V713
R02D3.3_a	ce	116	NELAKSVESQSKQVELQAKQKILQEDRNLEKLYQNLVATKLITPDDFWSDYYQKEGVSE	044499
R02D3.3_b	ce	231	CKEILKFTIQCEYLTRKISRSENYIQKKNLELVPHEMSEENFWKKFFQSHYFHR	044499
F2A19.20_a	at	82	LTPAEQLSMAEFELRFKLLRENSELQKLHKQFVESKVLTEDEFWSTRKKLLGKDS	Q9M322
F2A19.20_b	at	161	RTNRVTFNLTSEIIFQIFAEKPAVRQAFINYVPKKMTEKDFWTKYFRAEYLYS	Q9M322
SPAC16E8_a	sp	60	RVNSTNLEKDIDLQESLLTNNPDLLQTFKEAVMKGHLSNEQFWSTRLHLLRAHA	013745
SPAC16E8_b	sp	134	VDNQMKVSLTGQQIHDMFEQHPLLRKVYDKHVP-PLAEGEFWSRFFLSKLCKK	013745
B8B20.390_a	nc	147	WFEDDMLKADVELQQSLMKKDKALAHIYND[6]DSLSDASFNSQFWATRISLLRAYA	Q9P5N7
B8B20.390_b	nc	227	ENGELKLNINHEQVQLIFQQHPLVKRIYNENVP-KLTESEFWSRFFLSRLSKK	Q9P5N7
Hypo47.2	hs	146	WLSQFCLEEKKGEISELLVGSPSIRALYTKMVPAAVSHSEFWHRYFYKVHQLE	Q9NW68
Y97E10AR.6	ce	294	WISRFNLDEYDGEINILLANNPSLRQMFANLVPGSVNHETFWKRYFYAIEVAE	CE27417
F25G13.200	at	207	WSLGLKLEEKRNEIVELINGNKGVKEIYEEIVPVEVDAETFWRRYYYKVYKLE	Q9SV58
F15K9.5	at	179	WESAFSLDGKAEEMEKLLEENGDMKGVYKRVVPSMVDHETFWFRYFYRVNKLK	Q9ZVT6
HypoBAC	os	409	WRDAFRIDERKEEIEGVLKESPGLESFVERLVPSVVDYDMFWCRYFFAVDKLR	Q9LIX9
B23L21.150	nc	463	WVNEFDVDKKTEAIAADLDKYPELRATMEKLVPDQVPYADFWKRYYFLRHGIE	Q9P5L4
SPAC22A12	sp	167	WEKEISIDGKTEEISLLLEEYPDLRKQMESLVPSEVSYDDFWKRFFWHKEVVQ	013905
DOS2	sc	176	QLDPFDVDEKTEEICSILQGDKDISKLMNDIVPHKISYKDFWHIYFLQRNKIL	P54858
HypHS	hs	182	VQFNFDFDQMYPVALVMLQEDELLSKMRFALVPKLVKEEVFWRNYFYRVSLIK	AAH01468
SAP47	dm	272	VDFEFSYDTAYPTAIAIMAEDKALETMRFELVPKIITEENFWRNYFYRVSLII	Q24503
C16C2.4	ce	174	ANSEYTYEQQQAMATLLLKHDPNLANVRFQLVPKQVKENQFWQNYFYRIGLIR	017591
K7P8	at	86	NVKKDLSDWQEKHAVLVLSKSKELSQLRFKLCPRVLKEHQFWRIYFQLVRKIV	Q9LRX9
T16K5.150	at	195	FDDFEMTDAQYEHALAVENLASSLAALRIELCPAYMSEYCFWRIYFVLVHPIF	Q9M2X8
F20B24.15	at	227	IKNLEMSDAQRGHALAIERLAPRLAALRIELCPCHMSVGYFWKVYFVLLLSRL	Q9SGX8
HypOS	os	161	DENSIISDIQRDHMEAIEKLVPDLASLRARLCPSYMDIDVFWKIYFTLLESNL	Q9LWJ8
AT2G10950	at	137	DTEFELSEAQRAHASAIEDLVPGLVAVKNQVSSYMDDEHFWLIYFILLMPRL	Q9SKH9
T6L1.21	at	178	NVRKDLSEWQERHATLVLGSVKQISKLRYELCPRVMKERRFWRIYFTLVSTHV	Q9CAA2
F6H11.10	at	769	FSDFELADAQYEHALAVERLAPSLASLRIELCPEYMTENCFWRIYFVLVHPKL	049529
F20N2.15	at	424	STSSEQLSIKELELRFKLLRENRYRLHKQFVESKVLTEDEFWATRKKLLGKDS	Q9LFZ6
LMAJFV1	lm	340	WALHSLFDFDRDVQEGLLASA-EVRAHRYRLVPARLKEVTFWANYFWKVHCVG	060968
PFC1055W	pf	302	QKLSKSVEINNELRKLILCENKELKKLYDYYIENNILSDSKFWFFLFNNKYSHL	097305
Consensus (80%)			hp.phlhpl.hh.phss.hp.ppFW.haa.hh.	
Sec.str.pre	d.		hhнннннннннннннh, hнннннннhннннннннн	
				TiBS

Fig. 1. Multiple sequence alignment of BSD domains of BTF2-like transcription factors (TFB1, BTF2, TFB1dm, R02D3.3, F2A19.20, SPAC16E8, B8B20.390), DOS2-like proteins (Hypo47.2, Y97E10AR.6, F25G13.200, F15K9.5, HypoBAC, B23L21.150, SPAC22A12, DOS2), proteins related to a synapse-associated protein (HypHS, SAP47, C16C2.4, K7P8 T16K5.150, F20B24.15, HypOS, AT2G10950, T6L1.21 and, with an N-terminal Ubox, F6H11.10), BTB domain-containing protein (F20N2.15), and other hypothetical proteins (LMAJFV1, PFC1055W). First column: protein names (multiple domains in the same protein are labelled a and b); second column: species names; third column: start of the domain in the respective sequences: rightmost column: database accession numbers. Partially conserved (>50%) negatively charged residues are shown in red; conserved hydrophobic residues in blue; conserved aromatic residues in bold blue, and other conserved residues in bold black. The consensus sequence (conserved in 80% of the sequences) is shown below: h. p. s. l and a indicate hydrophobic, polar, small, aliphatic and aromatic residues, respectively. The predicted secondary structure taken from the consensus of the alignment (H, helix predicted with expected average accuracy >82%; h, helix predicted with expected average accuracy <82%) [9]. Several expressed sequence tags exist in various eukaryotes, supporting the idea that the BSD domain is found in a wide species range (data not shown). The domain boundaries shown have been predicted after analysis of all family members. For all subfamilies, PHD only predicts a coherent secondary structure in the region displayed here. Similarities beyond the region (if existing at all) are confined to individual subfamilies; other subfamilies contain deletions or segments of low complexity in regions preceding or succeeding the predicted boundaries [e.g. N terminus of T16K5.150 (185–199) or C terminus of DOS2 (240–265)]. Abbreviations: at, Arabidopsis thaliana; BSD, found in BTF2-like transcription factors, synapse-associated and DOS2-like proteins; BTB, Broad-complex, Tamtrack and Bric a Brac; ce, Caenorhabditis elegans; dm, Drosophila melanogaster; hs, Homo sapiens; Im, Leishmania major, nc, Neurospora crassa; os, Oryza sativa; pf, Paramecium falciparum; sc, Saccharomyces cerevisiae; sp, Saccharomyces pombe; Ubox, a modified Ring finger domain associated with ubiquitination.

protein [6]). The HMMer search reveals significant similarity between members of this family and DOS2-like proteins (E value = 7.2×10^{-7}). In further HMMer iterations (including identified homologues), the suspected homologies of synapse-associated proteins to BTF2-like transcription factors (E value = 2.3×10^{-3}), a BTB/POZ-domain-containing protein [7] and other hypothetical proteins in protozoans (Fig. 2), were confirmed.

We named the newly discovered region the BSD domain (after the <u>B</u>TF2-like transcription factors, <u>synapse-associated</u> and <u>D</u>OS2-like proteins in which it is found). A multiple sequence alignment was

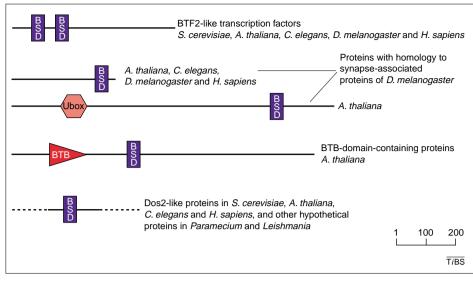


Fig. 2. Domain architecture of proteins containing the BSD domain. Only proteins with distinct modular organizations are shown. The domain names are according to those in the Simple Modular Architecture Research Tool [13] (http://smart.embl-heidelberg.de). Note that the name synapse-associated protein might be misleading as synaptic localization has so far only been demonstrated for *Drosophila melanogaster* [4], and close homologues are found in species without synapses. Abbreviations: BSD, found in <u>B</u>TF2-like transcription factors, <u>synapse-associated</u> and <u>D</u>OS2-like proteins; BTB, <u>B</u>road-complex, <u>T</u>amtrack and <u>B</u>ric a Brac; Ubox, a modified Ring finger domain associated with ubiquitination.

generated [8] for all candidates to identify the potential domain boundaries (Fig. 1). From this, the BSD domain appears to be ~60 amino acids in length. Secondary structure prediction with PHD [9] indicates the presence of three α helices, which probably form a three-helical bundle in small domains. The third predicted helix

Box 1. Artificial in silico support of function prediction*

Currently, one of the proteins in the alignment, DOS1, is annotated in most databases to be involved in single-copy DNA replication and ubiquitination. This assumed function would match our findings that the BSD domain is present in basal transcription factors and could have a role in DNA-binding.

The functional description is based on a mutation in a region around the yeast open reading frame YDR068W (dating back to 1995), and the gene was originally named DOS1 in *Saccharomyces cerevisiae*. Further studies revealed that the mutation is localized in an adjacent gene; therefore, YDR068w was renamed DOS2 in GenBank at a later stage, without functional description. However, in the meantime, the old name and functional implications of the gene were imported into different public databases, where the erroneous entry can often still be found (Table I).

Table I. Variable nomenclature found in databases

Date of information	Correct dat	abase entries	Erroneous database entries Wrong gene names: DOA4,DOS1, UBP4, SSV7, NPI2, YDR069C		Erroneous database entry Correct gene name, but functional prediction of DOS1	
retrieval	YD8554.01,	YD9609.22,				
September -October	GenBank:	AAA66522 NP 010353	Sptrembl:	O13905 Q9P5L4		
2001	EMBL:	CAB16584 (one representative)	SWISSPROT: EMBL:	P54858 CAB91683		
			PIR:	S54052 T49702		
28 November 2001	GenBank:	AAA66522 NP_010353	Sptrembl:	O13905, Q9P5L4	SWISSPROT: P54858	
	EMBL:	CAB16584 (one representative)	EMBL: PIR:	CAB91683 S54052 T49702		

*Tim Formosa (University of Utah, Salt Lake City, UT 84112-5330, USA) was a coauthor of Box 1.

contains neighbouring phenylalanine and tryptophan residues – less common amino acids that are invariant in all the BSD domains identified and that are the most striking sequence features of the domain (Fig. 1). The BSD domain is found in a variety of species from primal protozoan to human, indicating a conserved, probably important, function.

Although the BSD domain occurs in very different protein families (e.g. synapse-associated proteins, hypothetical proteins and transcription factors), the presence of the novel domain in transcription factors suggests a role in chromatin-associated processes. The domain architectures of additional BSD domain-containing proteins are consistent with this assumption, but also make other functions feasible. For example, the BSD domain can co-occur with Ubox domains (Fig. 2), which are known to be involved in ubiquitination [6]. Although several proteins involved in the ubiquitination process are known to be associated with chromatin [10,11], this is not a prerequisite. BSD domains can also precede a BTB domain (Fig. 2), a protein-protein interaction domain that frequently occurs in transcription factors, in which it is succeeded by ZnF C2H2 DNA-binding domains [7,12]. These findings suggest that the BSD domain could have a role in DNA binding, although it should be noted that neither synapseassociated proteins nor DOS2-like proteins are known to be associated with chromatin.

In summary, the delineation of the BSD domain and its boundaries (Fig. 1) should allow directed structural studies to test the involvement of this domain in chromatinassociated or more general processes.

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Sec61 β – a component of the archaeal protein secretory system

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Sec61p/SecYEG complexes mediate protein translocation across membranes and are present in both eukaryotes and bacteria. Whereas homologues of Sec61 α /SecY and Sec61 γ /SecE exist in archaea, identification of the third component (Sec61 β or SecG) has remained elusive. Using PSI-BLAST, the archaeal counterpart of Sec61 β has been detected. With the identification of the Sec61 β motif, functions for a universal family of archaeal proteins can be predicted and the archaeal translocon system can be definitively detected.

Detection and rationalization of motifs in membrane proteins are more difficult than in soluble proteins because of their biased amino acid composition that is restricted to mostly hydrophobic residues and to a limited number of available spatial structures. The single transmembranespanning protein Sec61_β (Sbh1p in yeast) interacts with two other integral membrane proteins (Sec 61α and Sec 61γ) to form the core of the eukaryotic protein translocation machinery (reviewed in Refs [1,2]). The bacterial counterpart of this machine consists of a similar complex (SecYEG), with SecY and SecE representing homologues of Sec61 a and Sec61_γ, respectively [3]. The third bacterial membrane protein, SecG, differs somewhat from Sec61 β in both the number of membrane-spanning regions and residue conservation. This divergence brings into question the evolutionary origins of this third subunit, although SecG and Sec61 $\!\beta$ both function to stimulate protein translocation activities and are thought to

be homologous [4,5]. Although archaeal homologues of SecY/Sec61 and SecE/Sec61y exist, the identification of an archaeal homologue to either Sec61 β or SecG has remained elusive. The Sec61p/SecYEG system is universally present in all eukaryotes and bacteria for which completely sequenced genomes are available (T. Cao and M.H. Saier, Jr, unpublished). Thus, the absence of a Sec61^β/SecG homologue in archaea is puzzling. Based on sequence analyses, we have identified the third component of the archaeal translocation machinery. The archaeal counterpart resembles eukaryotic Sec61 β , suggesting an overall functional similarity between the translocation apparatus of archaea and the eukaryotes. Although this functional similarity awaits experimental conformation, it mimics similarities displayed in other universal processes such as DNA replication, transcription and translation [6], and provides additional data for the studies of archaeal evolutionary origin [7].

We first detected a possible archaeal counterpart (gi | 15920503) to the human Sec61 β sequence (gi | 5803165) using PSI–BLAST [8] (parameters described in Fig. 1). Upon searching protein databases for related archaeal sequences, we found hits in all but two of the completely sequenced archaeal genomes. Searches against the nucleotide databases of these genomes suggested that these sequences (AE000914 and AE006662) were missed in gene prediction efforts. To substantiate the link to eukaryotic Sec61 β sequences, we generated a position-specific scoring matrix with a multiple sequence alignment

of the archaeal Sec61 β . Using this matrix, we initiated PSI–BLAST searches with each sequence from the alignment as a query. Two archaeal sequences used as queries (gi | 15920503 or RAP00437) identified the eukaryotic Sec61 β sequence (gi | 15239337) with significant statistics (E-value 0.002). This E-value, representing the estimated number of alignments with scores no less than that of a given alignment that is expected to occur in a database search by chance [8], falls below the threshold observed for distant homologues (E = 0.01) [9].

The short 45-residue motif identifying Sec61 β consists of a single, mostly hydrophobic stretch of ~20 amino acids preceded by a region of similar size that starts with several small amino acids and displays a particular residue conservation pattern (Fig. 1). The hydrophobic segment is predicted to form a transmembrane helix, with the C terminus of the helix defined by small and positively charged residues. The sequence of the helix incorporates a small residue at the beginning of the third turn and a relatively conserved histidine in the last turn. We suggest that the most conserved residue in the motif (proline) forms part of the N-terminal cap structure of this helix. The sequence between this helix and the stretch of small residues at the N terminus of the motif is characterized by four predominantly charged positions, having two negative charges surrounded by positive charges on either side. The archaeal sequences additionally contain conserved positively charged residues N-terminal to the transmembrane helix,